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Direct evidence for close proximity of catalytic and regulatory domains of heterodimeric sGC based on fluorescence resonance energy transfer

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Background

To examine the structural organisation of heterodimeric soluble guanylyl cyclase (sGC) fluorescence resonance energy transfer (FRET) was used to estimate distances between fluorescent proteins fused to the amino- and carboxy-terminal ends of the sGC β_1 and α subunits.

Methods

The FRET donor, CFP, and FRET acceptor, YFP, were fused to amino- and carboxy-terminal ends of sGC subunits. After generation of recombinant baculovirus strains fluorescent tagged sGC subunits were co-expressed in Sf9cells. Fluorescent variants of sGC were analyzed *in vitro* in cytosolic fractions by sensitized emission FRET. In addition, fluorescent tagged sGC subunits were analyzed *in vivo* using confocal laser scanning microscopy and fluorescence lifetime imaging (FLIM) on an inverted microscope.

Results

Carboxy-terminal fluorescent-tagged sGC combinations displayed NO stimulated sGC activity similar to the non-tagged sGC heterodimer and showed *in vitro* and *in vivo* FRET values significantly higher than the negative control. Co-expression of amino-terminally tagged sGC showed also FRET. However, the enzyme complexes showed only basal enzyme activity. Co-expression of carboxy-terminally tagged α subunit with amino-terminally tagged β_1 subunit yielded a basally active enzyme complex that showed FRET. Co-expression of the amino-terminally

tagged α subunit with the carboxy-terminally tagged β_1 subunit resulted an enzyme complex that showed NO stimulated activity and FRET.

Discussion

Based on the ability of an amino-terminal construct of the β_1 subunit (HNOX) to inhibit activity of an heterodimer consisting only of the catalytic domains ($\alpha_{cat}\beta_{cat}$), Winger and Marletta [1] have proposed a direct interaction of the amino-terminal region of β_1 with the catalytic domains. Our results provide direct evidence that all four subunit termini of the heterodimeric enzyme complex are in proximity to each other. This supports the concept that sGC is structurally organized in a way that allows for direct interaction of the amino-terminal (HNOX) domains of β_1 (and α) with the carboxy-terminal catalytic region.

References

 Winger JA, Marletta MA: Expression and characterization of the catalytic domains of soluble guanylate cyclase: interaction with the heme domain. Biochemistry 2005, 44:4083-90.