

Meeting abstract

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Characterization of different G protein coupling properties of CB₁ and CB₂ cannabinoid receptors and GPR55 receptor using BRET

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from 15th Scientific Symposium of the Austrian Pharmacological Society (APHAR) Joint meeting with the Hungarian Society of Experimental and Clinical Pharmacology (MFT) and the Slovenian Pharmacological Society (SDF)
Graz, Austria. 19-21 November 2009

Published: 12 November 2009

BMC Pharmacology 2009, 9(Suppl 2):A43 doi:10.1186/1471-2210-9-S2-A43

This abstract is available from: <http://www.biomedcentral.com/1471-2210/9/S2/A43>

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Background

CB₁ and CB₂ cannabinoid receptors are G protein-coupled receptors which have been described to couple mainly to the G_{i/o} subfamily of G proteins. However, in some cell types and upon stimulation with certain cannabinoid agonists, activation of other G protein subtypes has also been observed. GPR55 is an orphan G protein-coupled receptor which has been suggested to be a novel member of the cannabinoid receptor family.

Methods

In this study we wanted to characterize the G protein activation properties of the two known cannabinoid receptors and GPR55 following stimulation with different cannabinoid ligands, using bioluminescence resonance energy transfer (BRET). We monitored the activation of different G protein subtypes (G_o, G_q, G_s or G₁₂) using *Renilla* luciferase-tagged wild type or chimeric Gα_o subunits (i.e. Gα_o with the C-terminal 5 amino acids replaced with those of Gα_q, Gα_s or Gα₁₂, respectively) co-expressed with EYFP-tagged α₁α₁₁ subunit and the receptor in CHO cells.

Results

We found that CB₁ was able to activate all four subtypes of G proteins, with different pharmacokinetic properties, following stimulation by non-selective (WIN55 and 2-AG) or CB₁-selective (ACEA) cannabinoid agonists. Basal activity of CB₁ could also be detected with G_o and G₁₂ subtypes, as the CB₁ inverse agonist AM251 caused significant

BRET increase (i.e. G protein subunit association) when tested with these G proteins. In contrast, CB₂ showed no G protein activation other than G_o, upon either WIN55 or 2-AG stimuli. Stimulation of GPR55 with WIN55, 2-AG or AM251 did not alter the activity of the tested G proteins even at considerably high ligand concentrations.