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A role for cGMP-dependent protein kinase II in AMPA receptor trafficking and synaptic plasticity

Yafelle Serulle^{1,3}, Ipe Ninan^{2,4}, Daniela Puzzo^{2,5}, Maria McCarthy^{1,6}, Latika Khatri¹, Ottavio Arancio² and Edward Ziff*¹

Address: ¹Department of Biochemistry, NYU Langone Medical Center, New York NY 10016, USA, ²Department of Pathology and Taub Institute, Columbia University, New York, NY, 10032, USA, ³Department of Physiology, NYU Langone Medical Center, New York NY 10012, USA, ⁴Department of Psychiatry, NYU Langone Medical Center, New York NY 10012, USA, ⁵Department of Physiological Sciences, Viale A. Doria, 6, University of Catania 95125 Catania, Italy and ⁶Department of Psychiatry, Mt Sinai School of Medicine, New York NY 10029, USA

Email: Edward Ziff* - edward.ziff@nyumc.org

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Background

Trafficking of AMPA receptors (AMPARs) underlies the activity-dependent modification of synaptic strength and is regulated by specific interactions of AMPAR subunits with other proteins. We have reported (Serulle et al., 2007) [1] that the AMPAR subunit GluR1 binds the cGMP-dependent kinase type II (cGKII) adjacent to the kinase catalytic site, and that this interaction is increased by cGMP. In this complex, cGKII phosphorylates GluR1 at serine 845 (S845) leading to an increase of GluR1 on the plasma membrane. In neurons, cGMP is produced by soluble guanylate cyclase (sGC), which is activated by nitric oxide (NO), which is produced by nNOS under the control of the NMDA receptor.

Results

To distinguish the mechanism, we have measured the rate of exogenous GluR1 endocytosis in cultured primary neurons, either the wild type or the S845A or S845D mutants (Figure 1). We find that the S845A mutant (which cannot be phosphorylated) is endocytosed at rate of the wild type, while S845D (phosphomimetic) is endocytosed at a lower rate. Also, cGMP treatment, which elevates the endogenous GluR1 plasma membrane levels (Figure 2A), reduced the rate of of GluR1 endocytosis (Figure 2B).

Endocytosis of HA-GluR1 to Early Endosomes

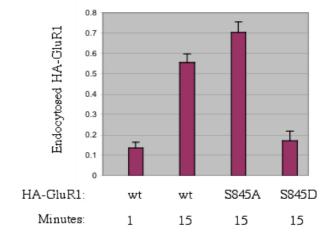


Figure I
Endocytosis of HAGIuRI is time dependent and is inhibited by the phosphomimetic mutation, S845D.

^{*} Corresponding author

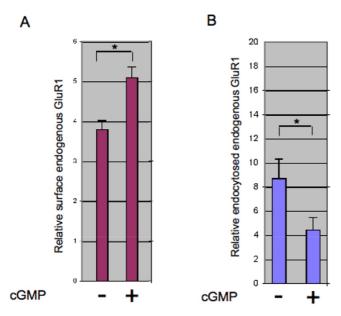


Figure 2
Elevation of surface GluRI and reduction of GluRI endocytosis by cGMP. A. cGMP, elevates surface GluRI.
B. cGMP decreases endocytosis of GluRI.

Conclusion

These data suggest that S845 phosphorylation increases the plasma membrane levels of GluR1 by reducing the rate of endocytosis.

References

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