

POSTER PRESENTATION

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Activation of cGMP-dependent protein kinase α and cAMP-dependent protein kinase A isoforms by cyclic nucleotides

Sabine Wolter^{1*}, Marina Golombek¹, Andreas Hammerschmidt¹, Frank Schwede², Hans-Gottfried Genieser², Roland Seifert¹

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Introduction

cAMP and cGMP are second messengers that play important roles in intracellular signal transduction of various external stimuli. Major functions of both are the activation of cAMP-dependent protein kinase A (PKA) and cGMP-dependent protein kinase G (PKG), respectively. PKA and PKG are members of the serine-threonine protein kinase superfamily and are involved in the control of various cellular processes.

PKA exists as an inactive tetramer of two regulatory (R) and two catalytic (C) subunits that dissociate in the presence of cAMP [1]. PKG is a polypeptide composed of a regulatory domain which contains two tandem cGMP-binding sites that interact allosterically and a catalytic domain [2]. Binding of cGMP to the regulatory domain increases the phosphotransferase activity of PKG.

Besides cAMP and cGMP, other cyclic nucleotides can tentatively function as second messengers. As shown by Desch et al. [3], the membrane-permeable cCMP-analogue dibutyryl-cCMP induces smooth muscle relaxation and activates PKGI in aortic tissue lysates by using the cGMP signal transduction pathway. Therefore, we have searched for further binding proteins by using cCMP-agaroses and cCMP-capture compounds, and we have identified the regulatory subunits of PKA as cCMP-interacting proteins.

Methods

PKA kinase activity with the regulatory subunits RI α and RII α of PKA and PKGI α kinase activity were

measured by *in-vitro* kinase assays in the presence of different cyclic nucleotides and analogues.

Results and discussion

We discovered that besides the known activators cAMP and cGMP, also all other cyclic nucleotides (cNMPs) studied (cCMP, cIMP, cUMP and cXMP) activate RI α and RII α , but with distinct activation constants (EC_{50}), different Hill slopes and different maximal effects (E_{max}). The most potent activator for RI α and RII α was cAMP. For RI α the Hill slopes indicated positive cooperativity for binding of all cNMPs, most notably for cAMP and cIMP. For RI α the most efficacious nucleotide was cIMP; for RII α the most effective activator was cUMP.

For PKGI α the most potent and effective activator was cGMP. All other cNMPs (cAMP, cCMP and cUMP) studied could also activate the enzyme, but the EC_{50} values and efficacies were much lower. For PKGI α we found a higher specificity for the cognate cNMP in comparison to PKA RI α and RII α .

By using membrane-permeable butyryl-cNMPs as activators, we found that only the mono-butyrylated cNMPs activated RI α and RII α of PKA. Monobutyryl (mb)-cCMP was a more potent activator for RI α and RII α in comparison to cCMP with a higher maximal activity for RI α . The Hill slopes for the activation of RI α with cCMP and mb-cCMP showed positive cooperativity.

To achieve substrate-specificity, A-kinase-anchoring proteins (AKAPs) bind to the regulatory subunits. In further studies the influence of different cNMPs on AKAP binding will be determined.

* Correspondence: Wolter.sabine@mh-hannover.de

¹Institute of Pharmacology, Hannover Medical School, Germany
Full list of author information is available at the end of the article

Table 1 LogEC₅₀ and E_{max} values for the activation of RI α (a), RII α (b) and PKGI α (c) with cNMPs

A						
	cAMP	cIMP	cGMP	cCMP	cUMP	cXMP
LogEC ₅₀ [M]	-7.1	-5.9	-4.9	-4.4	-4.2	-3.5
E _{max}	1.00	1.09	1.06	1.04	1.04	1.00
B						
	cAMP	cIMP	cGMP	cCMP	cUMP	cXMP
LogEC ₅₀ [M]	-7.0	-5.4	-4.5	-4.0	-4.4	-3.2
E _{max}	1.00	1.07	1.07	1.23	1.26	1.07
C						
	cGMP	cAMP	cCMP	cUMP		
LogEC ₅₀ [M]	-7.0	-5.0	-4.9	-4.1		
E _{max}	1.00	0.46	0.56	0.71		

Our data point to distinct cNMP-specific active conformation of RI α and RII α , the biological relevance of which has to be determined in future experiments.

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Author details

¹Institute of Pharmacology, Hannover Medical School, Germany. ²Biolog Life Science Institute, Bremen, Germany.

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References

1. Rehm H, Wittinghofer A, Bos JL: **Capturing cyclic nucleotides in action: snapshots from crystallographic studies.** *Nat Rev Mol Cell Biol* 2007, **1**:63-73.
2. Hofmann F: **The biology of cyclic GMP-dependent protein kinases.** *J Biol Chem* 2005, **280**:1-4.
3. Desch M, Schinner E, Kees F, Hofmann F, Seifert R, Schlossmann J: **Cyclic cytidine 3',5'-monophosphate (cCMP) signals via cGMP kinase I.** *FEBS Letters* 2010, **584**:3979-3984.

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